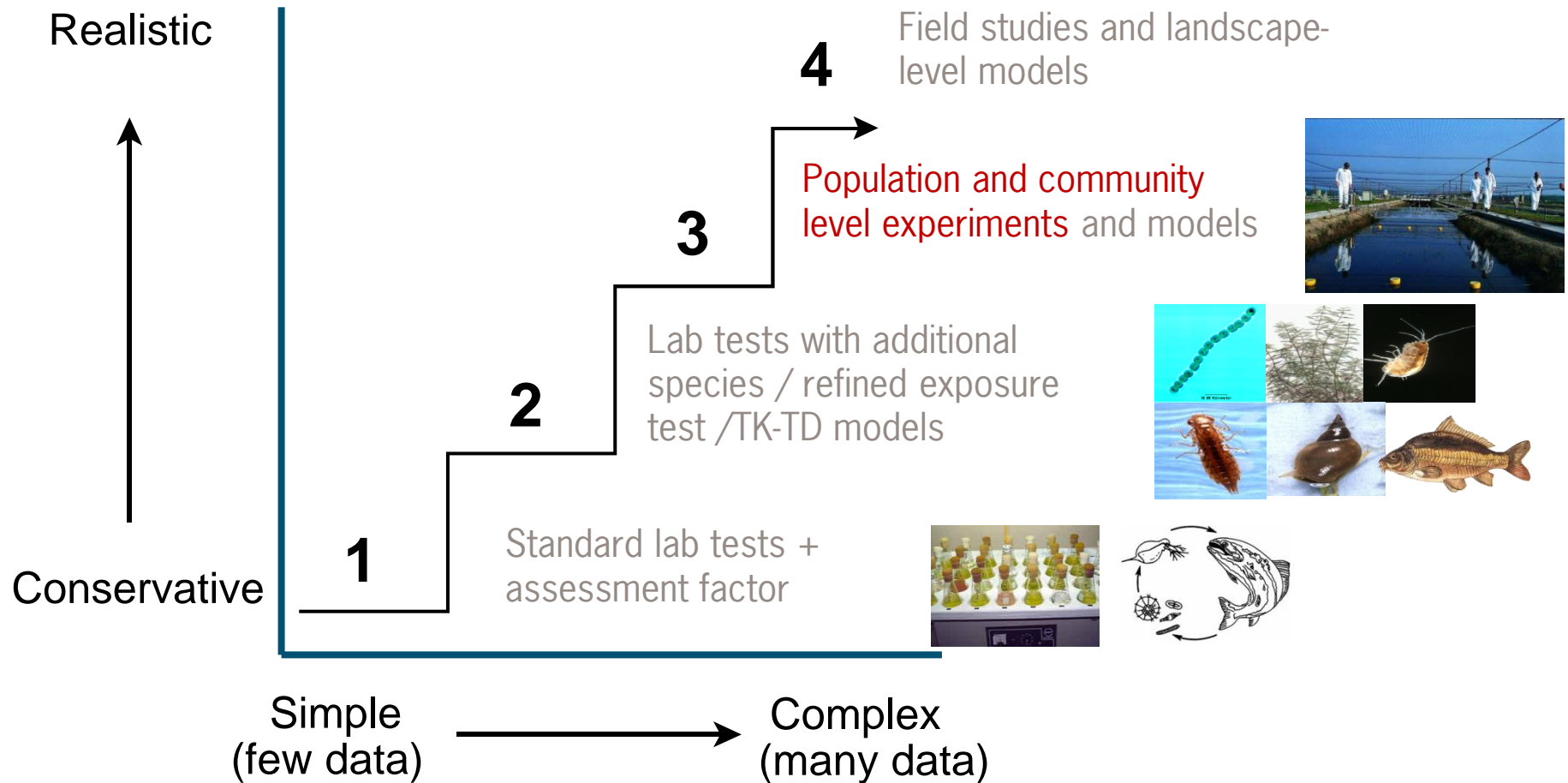


Aquatic Tier-3 effect assessment procedure (microcosm and mesocosm experiments)

Theo C.M. Brock



Tiered approach in effect assessment



Model ecosystems (microcosms, mesocosms)

- Constructed artificially with samples from, or portions of, natural ecosystems
- Housed in artificial containers or enclosures
- Reduction in size and complexity when compared with natural ecosystems
- Include an assemblage of organisms representing several trophic levels
- Community should be “adapted” to and in “equilibrium” with its ambient environment

Guidance on interpretation of micro/mesocosm tests: CLASSIC workshop (Giddings et al. 2002); RIVM report (De Jong et al. 2008); EFSA Aquatic Guidance Document (EFSA PPR 2013); Brock et al. 2014

Examples of microcosms and mesocosms



Most experience with lentic
freshwater ecosystems

MODEL ECOSYSTEMS versus NATURAL ECOSYSTEMS

PROS

- Experimental control and replication
- Their flexibility make them useful to a wide range of experiments
- More easily sampled and quantified
- Costs may be lower
- Less ethical to perform tests in natural ecosystems

CONS

- No perfect analogs of natural ecosystems
- Lack of large predators
- Wall effects (more periphyton, less turbulence)

Cause-and-effect relations in micro/mesocosms

Direct or primary effects

- Toxicological effects on survival, growth and/or reproduction of organisms
- Fairly well predictable on basis of laboratory toxicity tests and knowledge on field exposure

Indirect or secondary effect

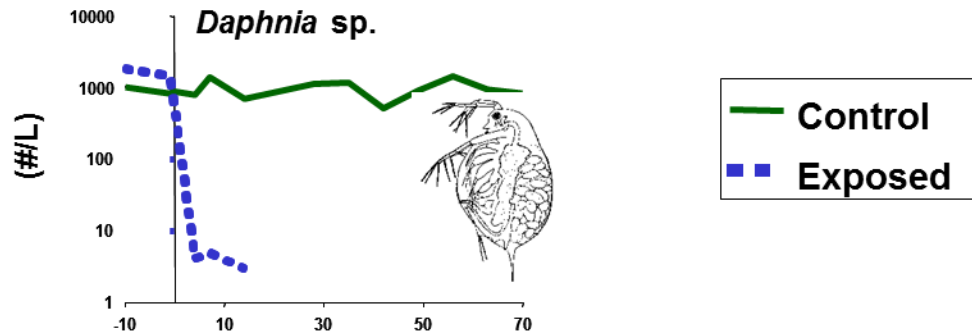
- Ecosystem changes that result from a reduction of (toxicant) susceptible species
- The same pesticide, dosed at a similar rate, may cause different secondary effects in different types of ecosystem

Delayed effects and biological recovery

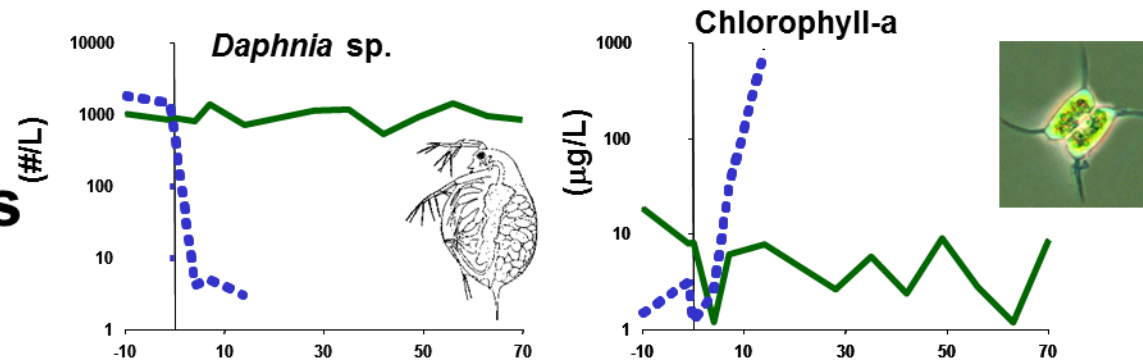
- The duration of the studies may be long enough to observe latency of effects and recovery of affected populations

Cause-and-effect relationships in micro/mesocosms

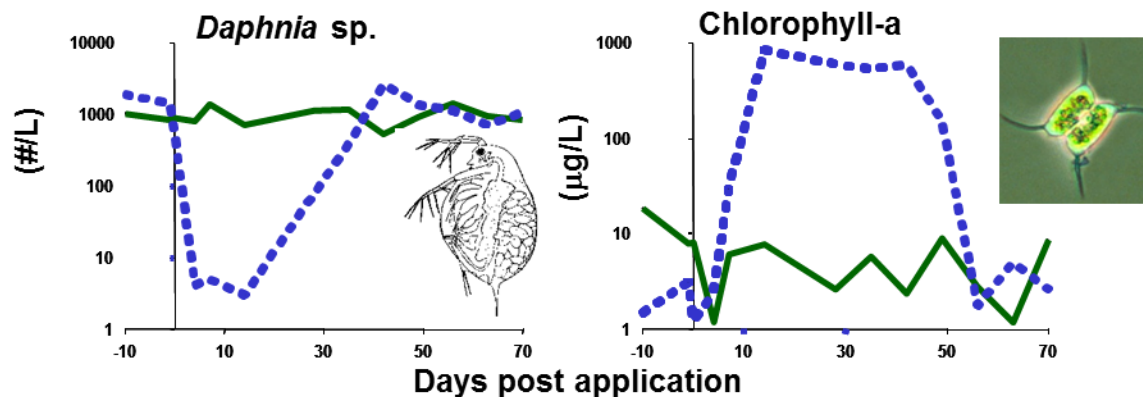
Sensitivity



Ecosystem interactions



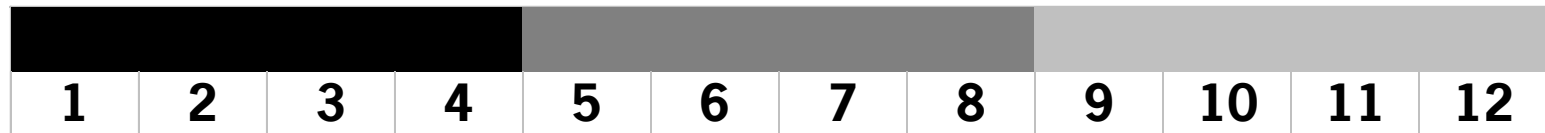
Recovery



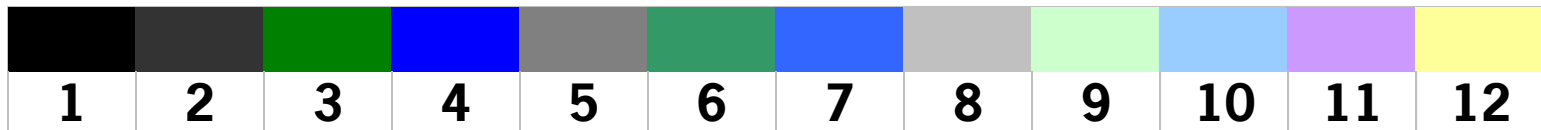
The conduct of micro/mesocosm tests

■ Experimental set-up: ANOVA versus Regression

- 4 replicates * 3 treatments



- 1 replicates * 12 treatments



Experimental set-up of micro/mesocosm tests

Combine the best of two worlds, replicated regression design

- E.g. the 3 replicates * 5 treatments + 4 controls experimental set-up
- Analysed with Williams test (combination between ANOVA and regression)
- Test assumes an increasing effect with an increasing dose

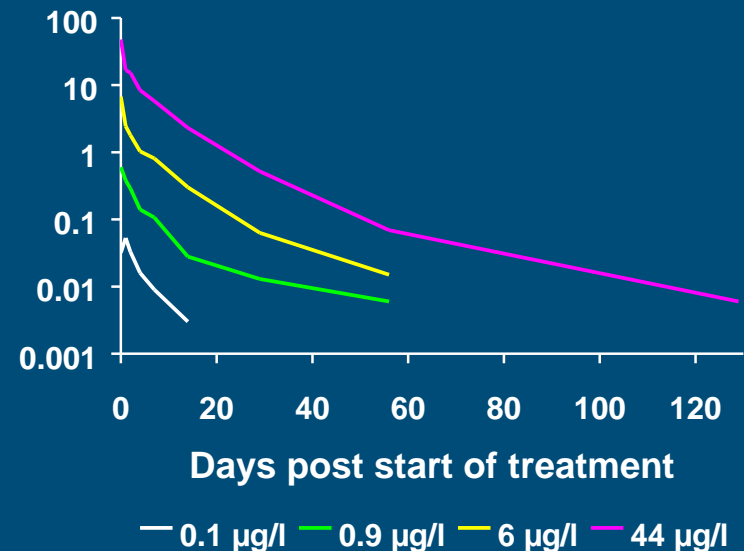
Chlorpyrifos: mesocosm experiment in the Netherlands



Single application (spray drift)

Macrophyte-dominated ditches

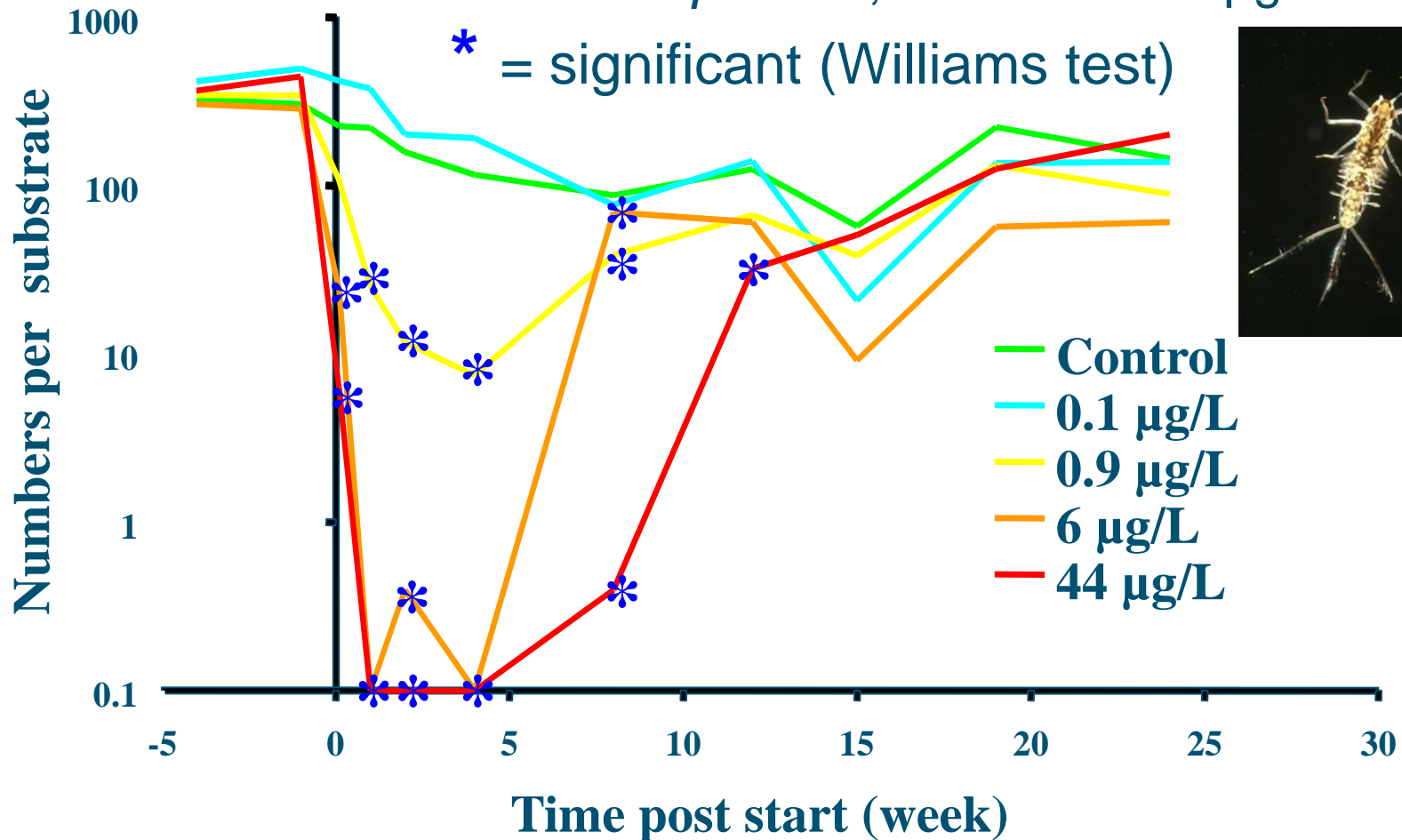
- 4 mesocosms controls
- 2 0.1 $\mu\text{g/L}$
- 2 0.9 $\mu\text{g/L}$
- 2 6 $\mu\text{g/L}$
- 2 44 $\mu\text{g/L}$



Univariate statistics sensitive endpoints

Cloeon dipterum; NOEC = 0.1 $\mu\text{g/L}$

* = significant (Williams test)



Allows to evaluate effects on more or less dominant populations

Chlorpyrifos example: Evaluation with univariate statistics

Typical outcome univariate NOEC calculations:

Results 19 of the 189 species collected

Taxon	Effect	-4	-1	0.1	1	2	4	8	12	15	19	24
Annelida												
<i>Oligochaeta</i>	+	>	>	>	>	>	0.9	6	>	>	>	0.9
<i>Stylaria lacustris</i>	+	>	>	>	>	>	0.9	0.9	>	>	>	>
Arthropods												
Crustacea												
<i>Simocephalus vetulus</i>	-	>	>	0.9	0.9	0.9	0.9	6	>	>	>	>
<i>Daphnia galeata</i>	-	n.p.	n.p.	n.p.	>	>	0.1	0.1	6	6	>	>
Ostracoda	-	n.p.	L!	6	0.9	6	6	0.9	6	6	6	>
Copepoda (mature stages)	-	>	>	>	L!	6	>	0.9	>	>	>	>
Copepoda (nauplii)	-	>	>	0.9	0.9	0.9	0.9	6	>	n.p.	>	n.p.
<i>Gammarus pulex</i>	-	n.p.	n.p.	n.p.	n.p.	n.p.	n.p.	0.1	L!	0.1	0.1	0.1
Insecta												
<i>Caenis horaria</i>	-	>	>	6	0.9	0.9	0.1	0.9	6	0.9	6	0.9
<i>Caenis luctuosa</i>	-	n.p.	n.p.	6	n.p.	0.9	0.1	0.9	>	>	n.p.	n.p.
<i>Cloeon dipterum</i>	-	>	>	0.9	0.1	0.1	0.1	6	>	>	>	>
Coenagrionidae	-	>	>	>	6	6	6	>	>	>	>	>
<i>Sialis lutaria</i>	-	n.p.	n.p.	n.p.	n.p.	n.p.	n.p.	0.1	n.p.	0.1	0.1	n.p.
<i>Hygrotus versicolor</i>	-	n.p.	n.p.	>	0.9	0.9	n.p.	6	0.9	n.p.	n.p.	n.p.
<i>Mystacides longicornis/nigra</i>		>	>	0.9	0.9	L!	0.1	>	6	6	6	>
<i>Ablabesmyia phatta/monilis</i>		>	>	6	0.9	0.9	0.9	6	6	>	0.9	n.p.
Ceratopogonidae	-	>	>	>	0.9	>	L!	6	6	>	>	>
<i>Chaoborus obscuripes</i>	-	L!	>	0.1	0.9	0.1	0.9	6	6	6	>	6
<i>Chironomus</i>	-	n.p.	n.p.	n.p.	>	n.p.	>	>	6	6	>	>

Chlorpyrifos example: Multivariate analysis



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PRINCIPAL RESPONSE CURVES: ANALYSIS OF TIME-DEPENDENT MULTIVARIATE RESPONSES OF BIOLOGICAL COMMUNITY TO STRESS

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†DLO Winand Staring Centre for Integrated Land, Soil and Water Research, P.O. Box 125, 6700 AC Wageningen, The Netherlands

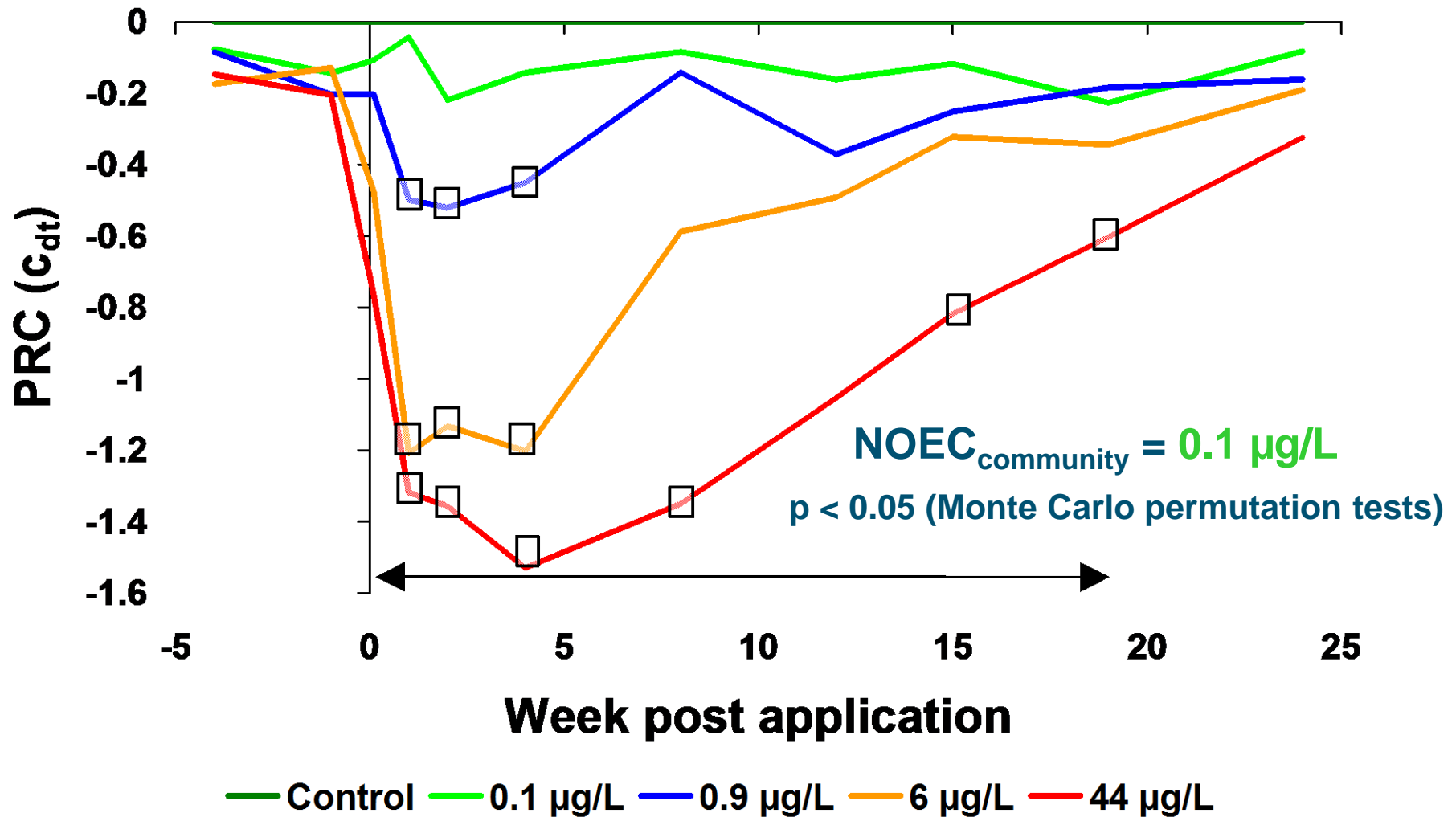
‡Centre for Biometry Wageningen, CPRO-DLO P.O. Box 16, 6700 AA Wageningen, The Netherlands

(Received 30 April 1997; Accepted 18 December 1997)

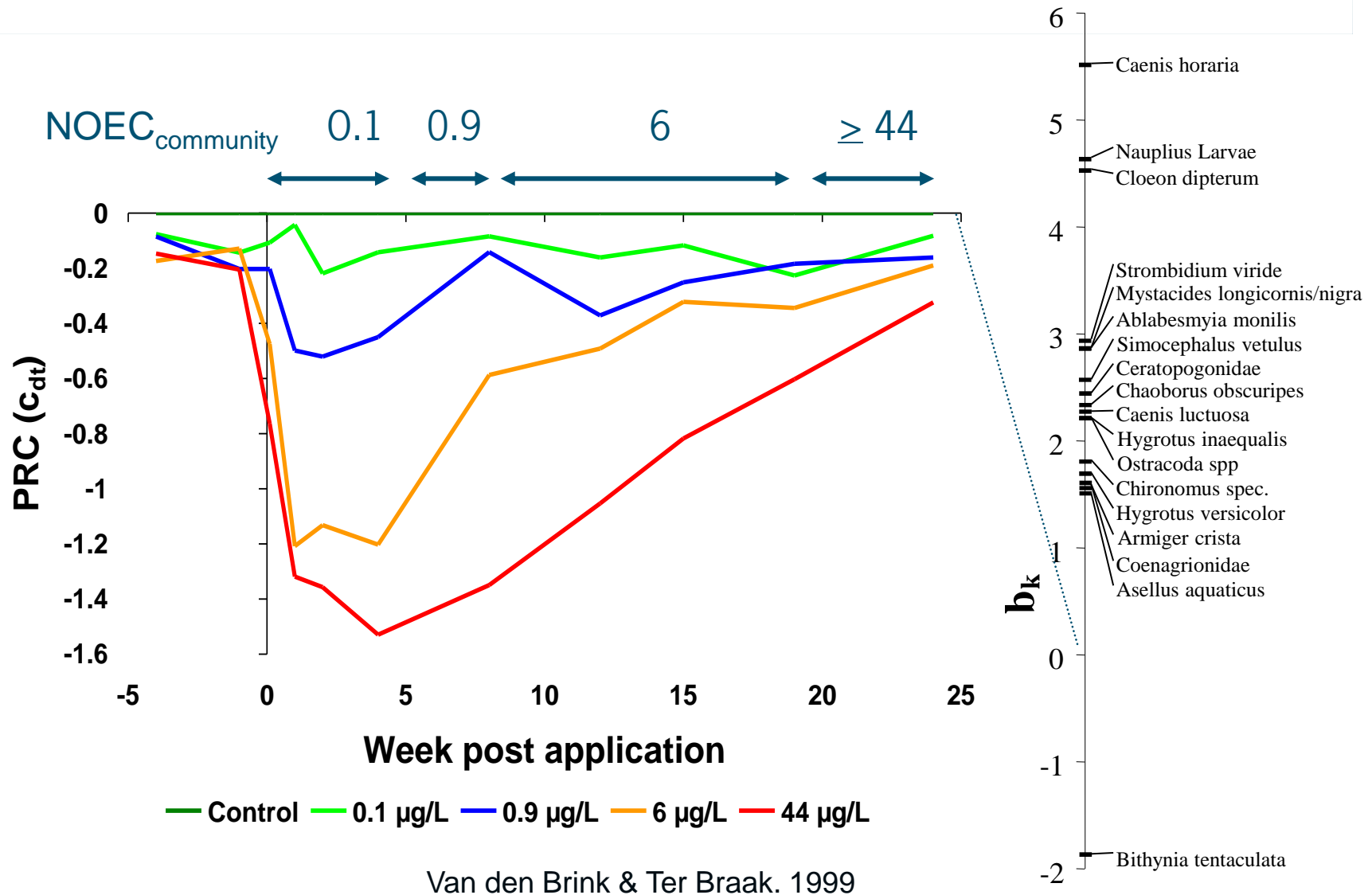
Abstract—In this paper a novel multivariate method is proposed for the analysis of community response data from designed experiments repeatedly sampled in time. The long-term effects of the insecticide chlorpyrifos on the invertebrate community and the dissolved oxygen (DO)–pH–alkalinity–conductivity syndrome, in outdoor experimental ditches, are used as example data. The new method, which we have named the principal response curve method (PRC), is based on redundancy analysis (RDA), adjusted for overall changes in community response over time, as observed in control test systems. This allows the method to focus on the time-dependent treatment effects. The principal component is plotted against time, yielding a principal response curve of the community for each treatment. The PRC method distills the complexity of time-dependent, community-level effects of pollutants into a graphic form that can be appreciated more readily than the results of other currently available multivariate techniques. The PRC method also enables a quantitative interpretation of effects towards the species level.

Keywords—Redundancy analysis Mesocosms Multivariate ordination techniques Pesticides Principal response curves

Chlorpyrifos example: Principal Response Curves



Chlorpyrifos example: Principal Response Curves



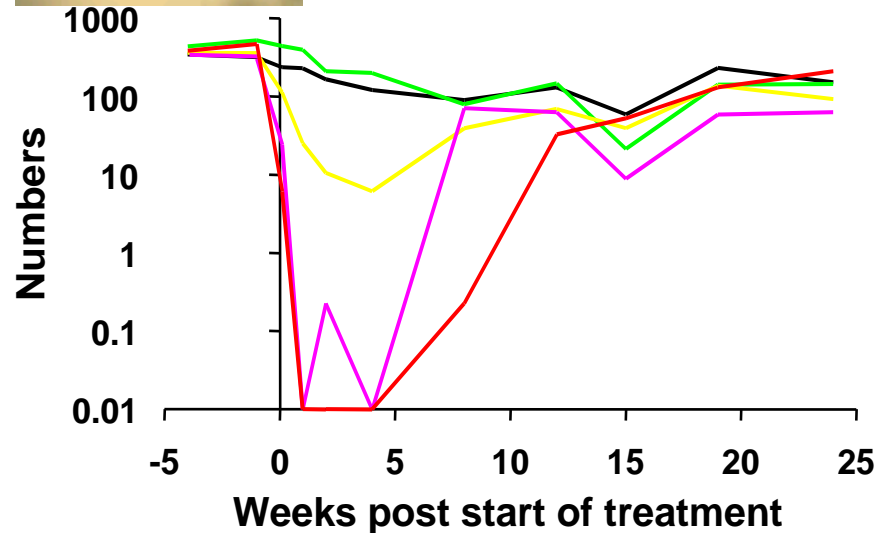
Allows to evaluate community-level effects

Chlorpyrifos exp.: sensitivity and recovery



Two mayfly species

Cloeon dipterum
NOEC = 0.1 µg/L



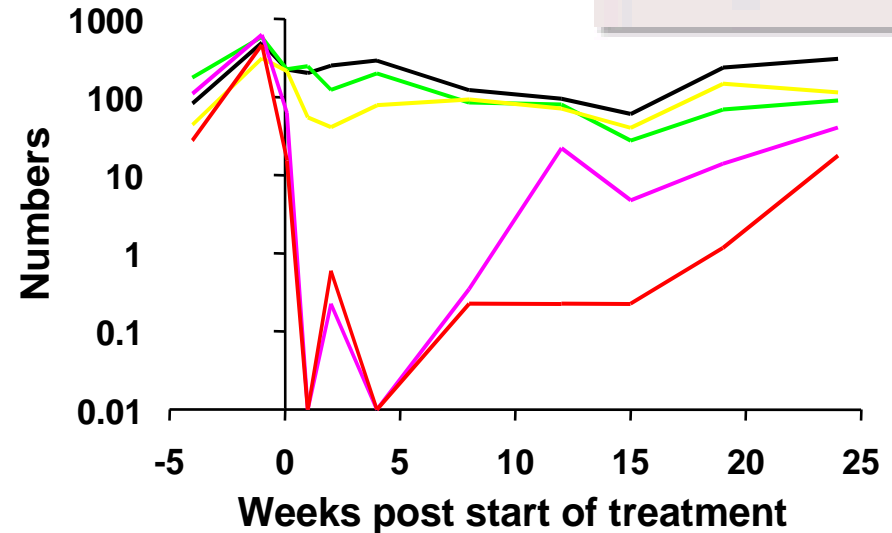
— Control — 0.1 µg/L — 0.9 µg/L — 6 µg/L — 44 µg/L

Relatively fast recovery

- appr. 3 generations/year
- egg deposition by terrestrial adults



Caenis horaria
NOEC = 0.1 µg/L



— Control — 0.1 µg/L — 0.9 µg/L — 6 µg/L — 44 µg/L

Relatively slow recovery

- 1 to 2 generations/year
- egg deposition by terrestrial adults

Rate of recovery is affected by number of generations per year

Chlorpyrifos example: Summary mesocosm experiment

Univariate statistics:

- Consistent NOECs (consecutive sampling dates) most sensitive populations $< 0.1 \mu\text{g/L}$ (slight short term effect)

Multivariate analysis

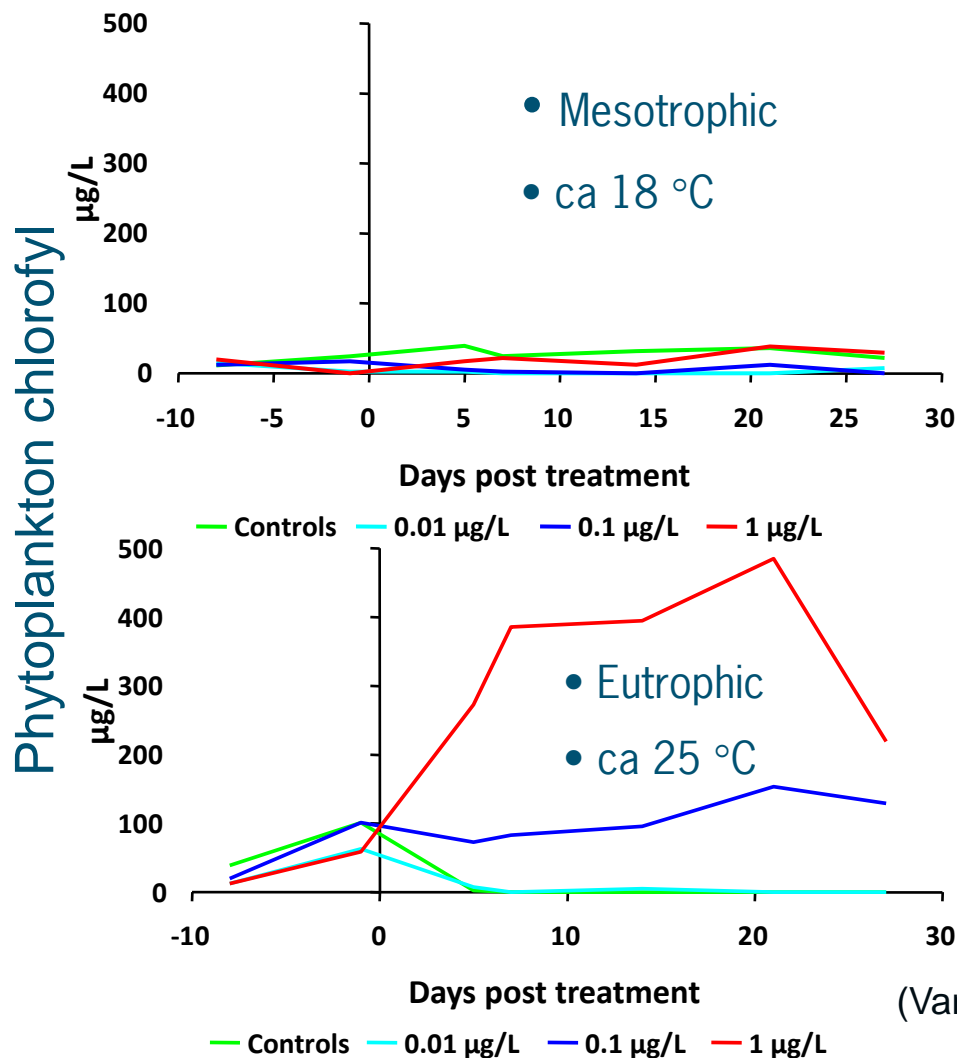
- NOEC invertebrate community = $0.1 \mu\text{g/L}$
- At treatment of $0.9 \mu\text{g/L}$ recovery within 8 weeks

Chlorpyrifos: semi-field experiments, single application exposure

Application regime	Class 1 NOEC	Class 2 LOEC	Class 3A LOEC	Type of test system	Location Reference
6 h pulse	0.1 µg/L	-	(5.0 µg/L)	Experimental streams	Australia Pusey et al. 1994
Single	0.1 µg/L	0.3 µg/L	1.0 µg/L	Outdoor microcosms	USA, Kansas Biever et al. 1994
Single	-	0.1 µg/L	<0.9 µg/L	Experimental ditches	Netherlands Van den Brink et al. 1996
Single	0.1 µg/L	-	1.0 µg/L	Lab microcosms Cool, Mesotr.	Netherlands Van Wijngaarden et al. 2005
Single	0.1 µg/L	-	≤1.0 µg/L	Lab microcosms Warm, Eutrophic	Netherlands Van Wijngaarden et al. 2005
Single	0.1 µg/L	-	1.0 µg/L	Outdoor mesocosms	Spain López-Mancisidor et al. 2005
Single	0.1 µg/L	-	1.0 µg/L	Outdoor microcosms	Thailand Daam et al. 2008
Single	-	-	0.5 µg/L	Pond enclosures	USA, Minnesota Siefert et al. 1989

Threshold levels for effects can be extrapolated with lower uncertainty than responses caused by higher exposures (but representatives of sensitive taxonomic groups need to be present).

Ecosystem interactions and indirect effects are context dependent



- 1 $\mu\text{g/L}$ chlorpyrifos
- reduction Cladocera (direct effect)
 - no algal bloom

- 1 $\mu\text{g/L}$ chlorpyrifos
- reduction Cladocera (direct effect)
 - pronounced algal bloom (indirect effect)

(Van Wijngaarden et al. 2005: *Pest Man Sci* 61:923-935)

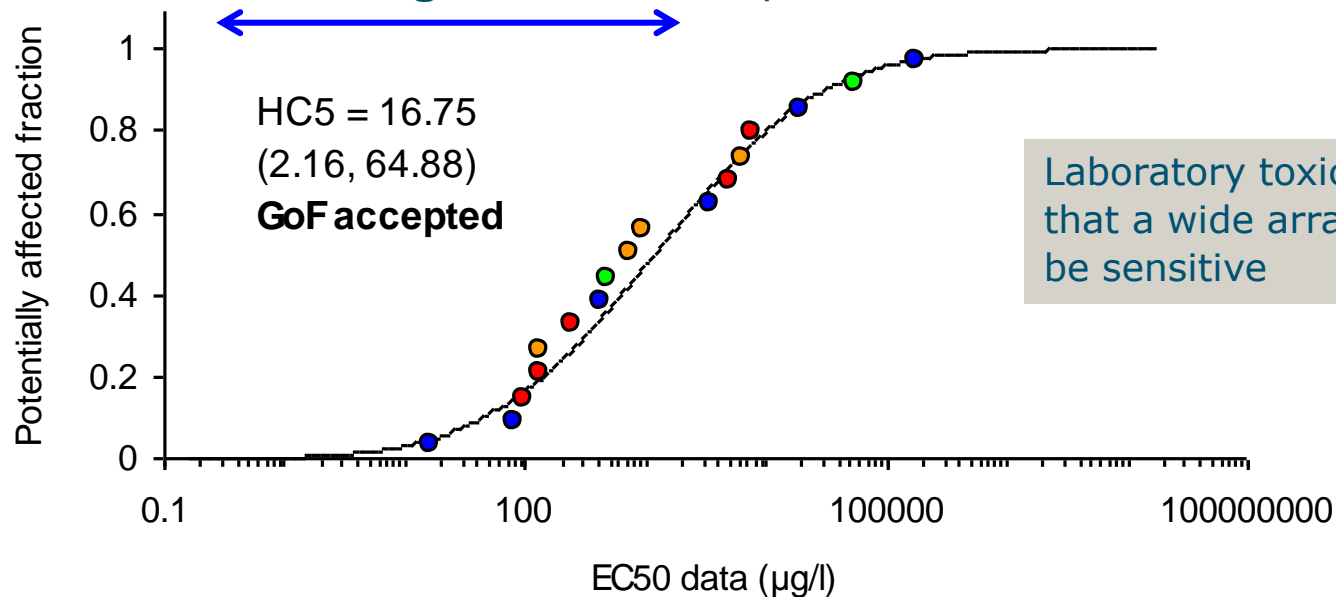
Magnitude and duration of direct and indirect effects is context dependent

(In)direct effects fungicide carbendazim



- Lentic macrophyte-dominated microcosms
- 0 – 3.3 – 33 – 100 – 330 – 1000 $\mu\text{g/L}$
- chronic exposure regime (constant during 4 weeks)

Concentration range microcosm experiment

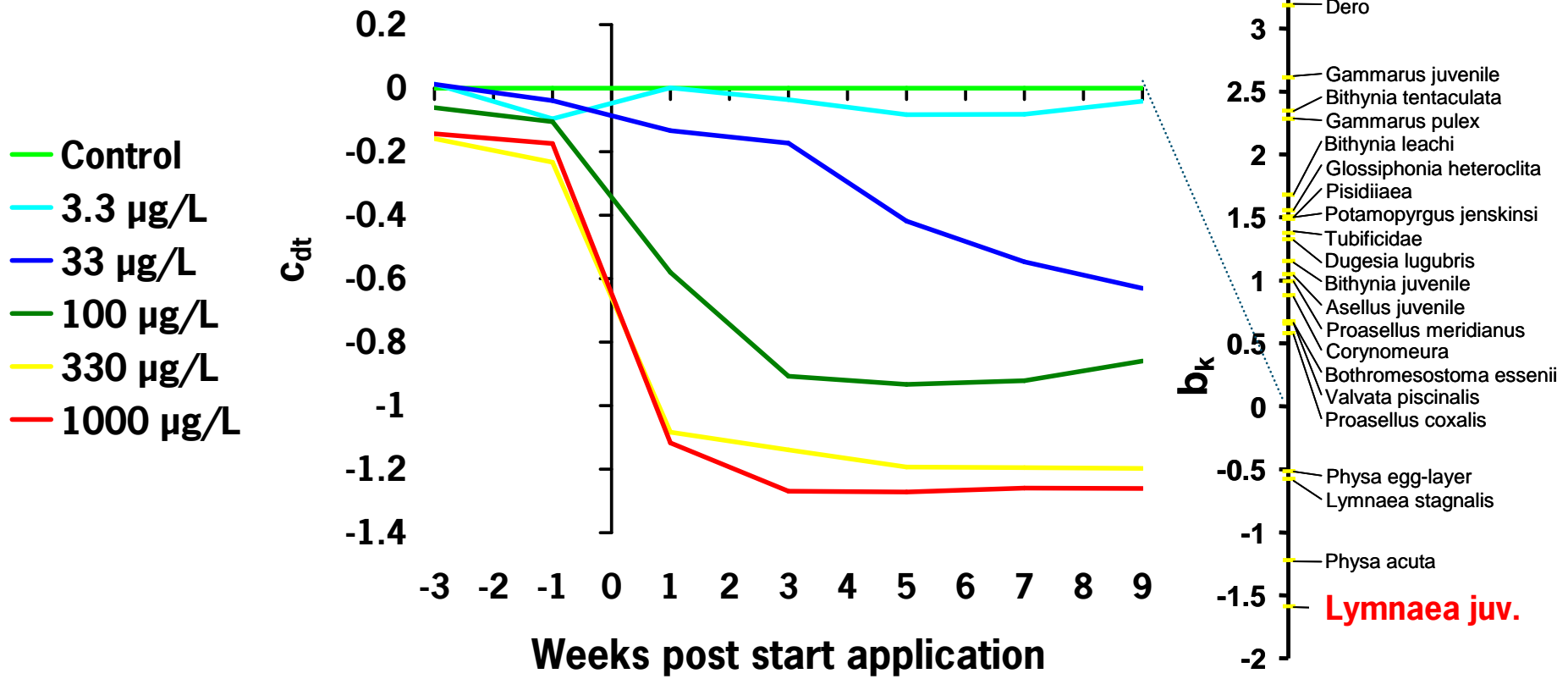


Laboratory toxicity data suggest that a wide array of species may be sensitive

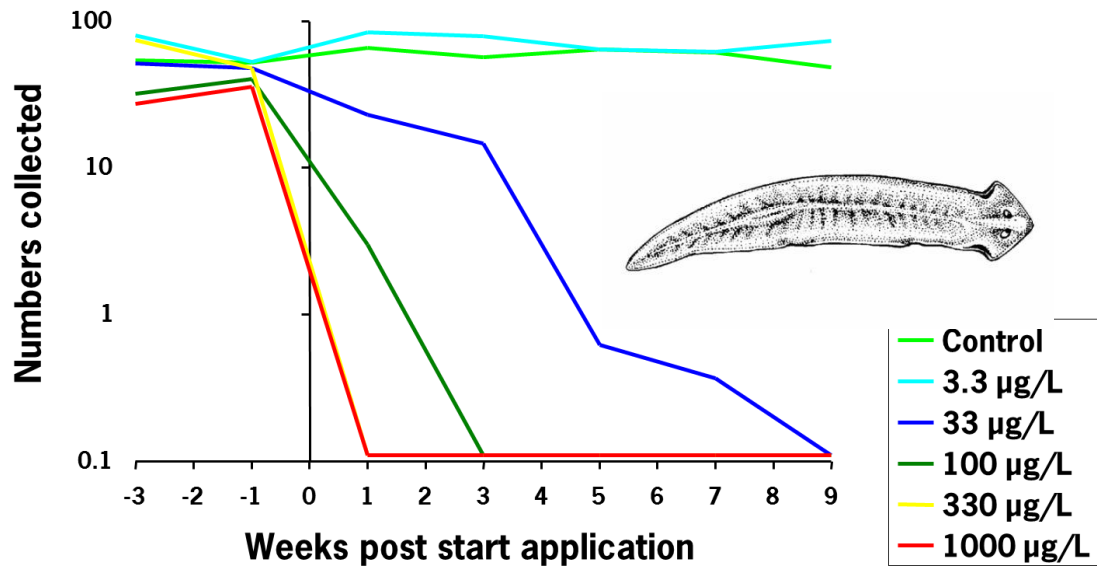
Carbendazim microcosm experiment

Multivariate analysis (PRC method)

NOEC = 3.3 $\mu\text{g/L}$



(In)direct effects fungicide carbendazim

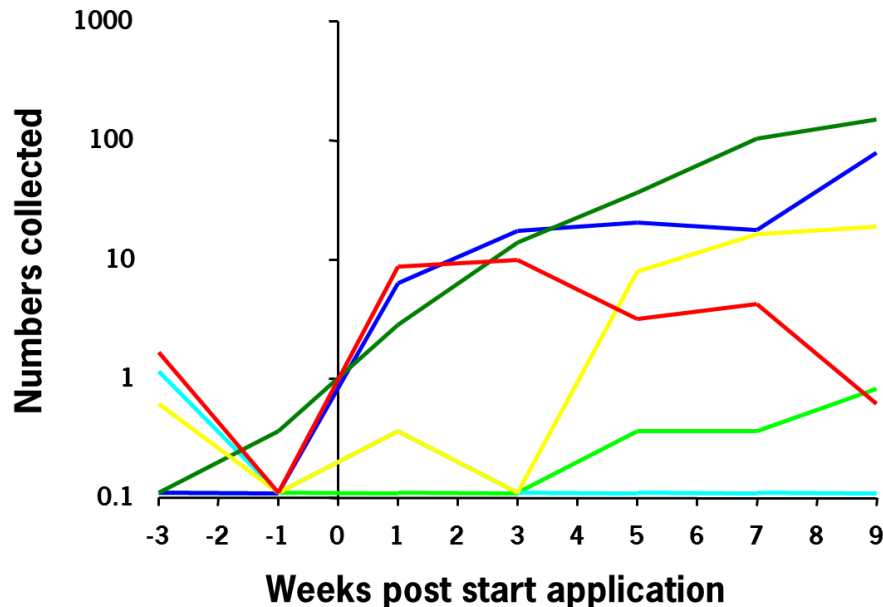


Dugesia tigrina

- Flatworm
- Direct effect
- NOEC = 3.3 µg/L
- Concentration – time dependency

Lymnaea stagnalis

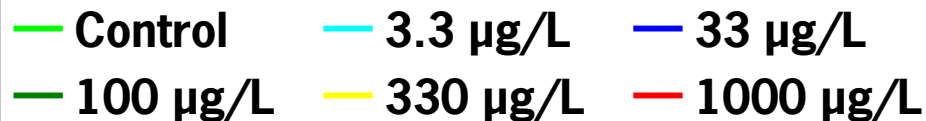
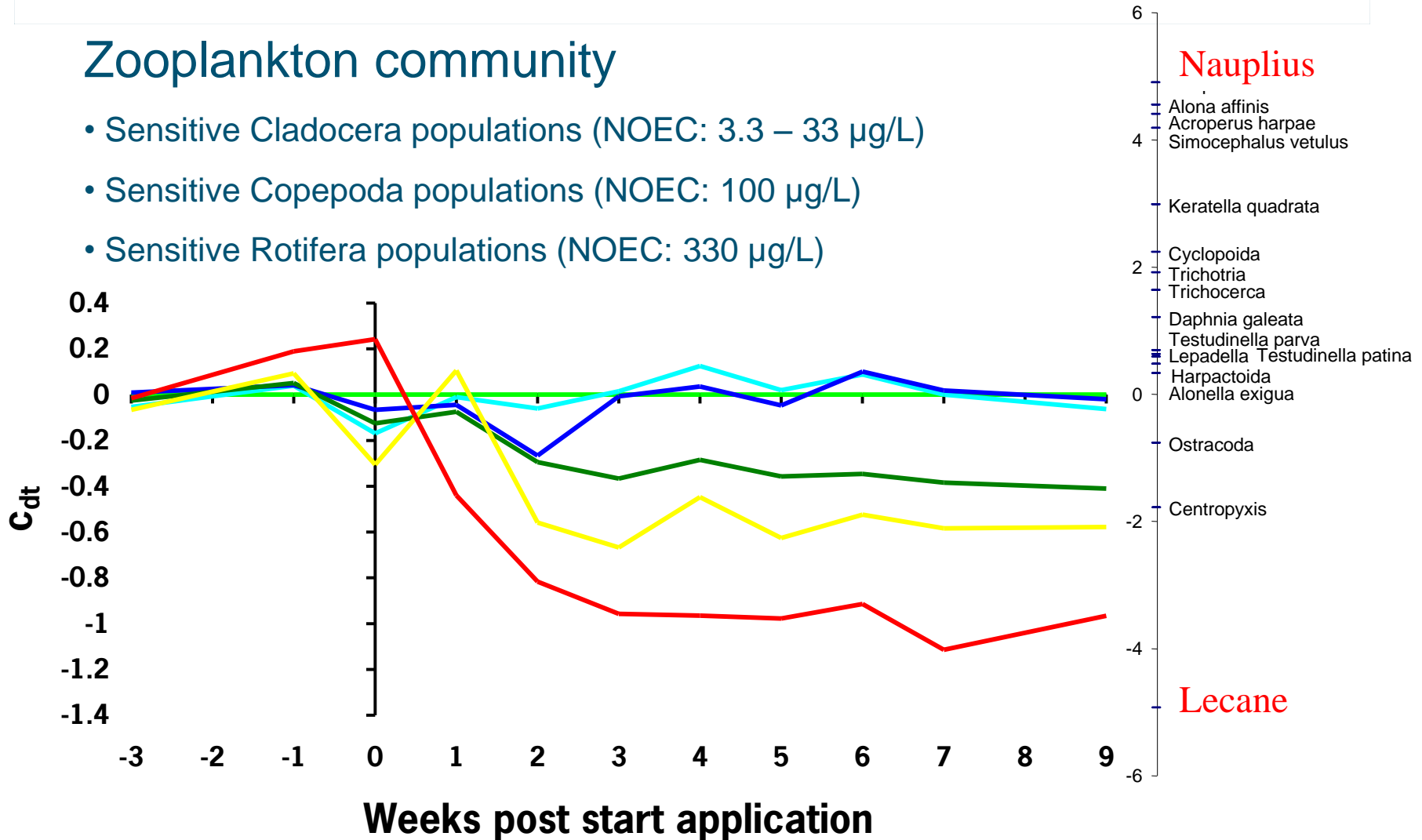
- Snail
- Indirect effect
- NOEC = 3.3 µg/L
- Less clear concentration – time dependency



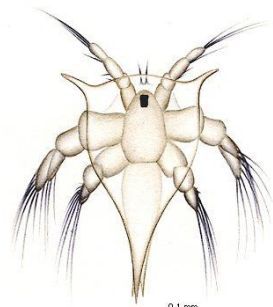
Carbendazim microcosm experiment

Zooplankton community

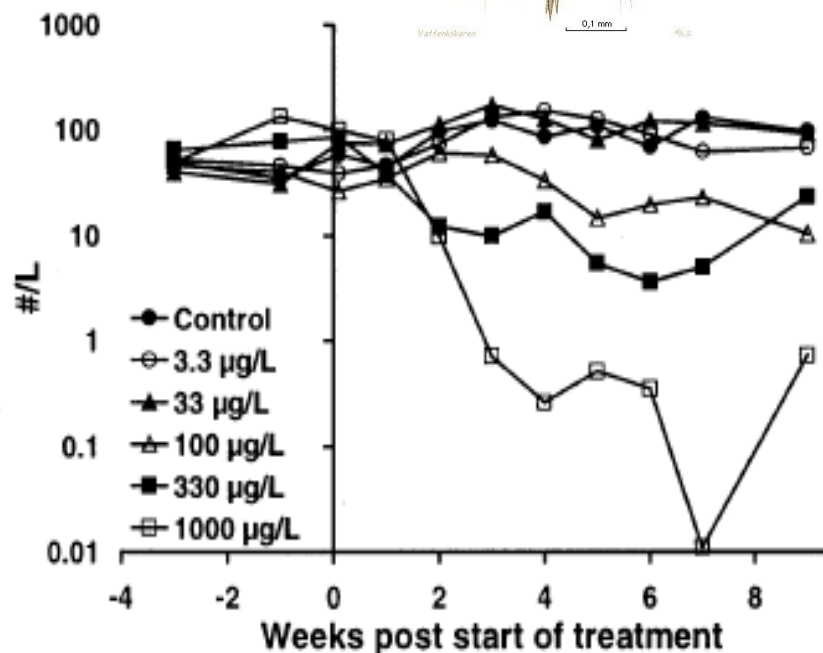
- Sensitive Cladocera populations (NOEC: 3.3 – 33 $\mu\text{g/L}$)
- Sensitive Copepoda populations (NOEC: 100 $\mu\text{g/L}$)
- Sensitive Rotifera populations (NOEC: 330 $\mu\text{g/L}$)



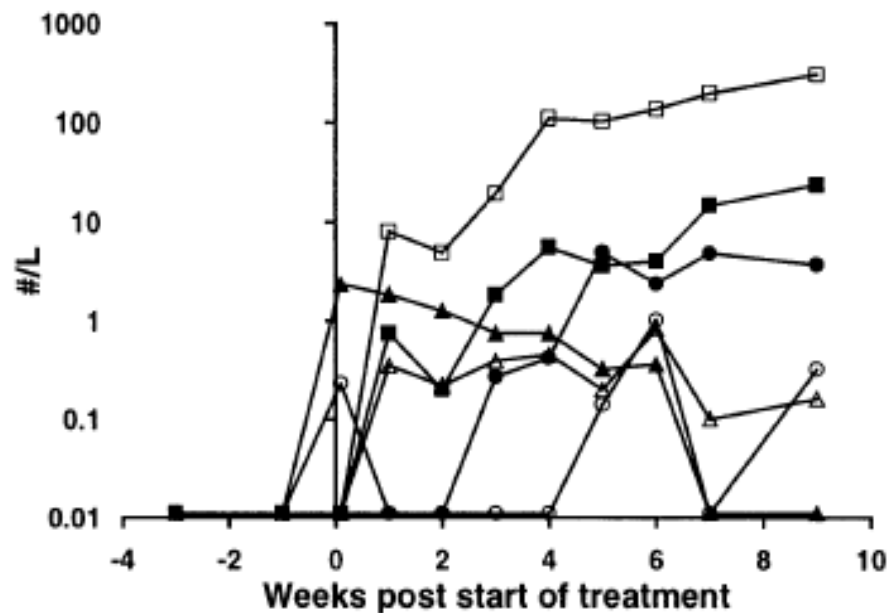
(In)direct effects fungicide carbendazim



Nauplius 0,1 mm



Nauplius (NOEC = 33 µg/L)
Direct effect

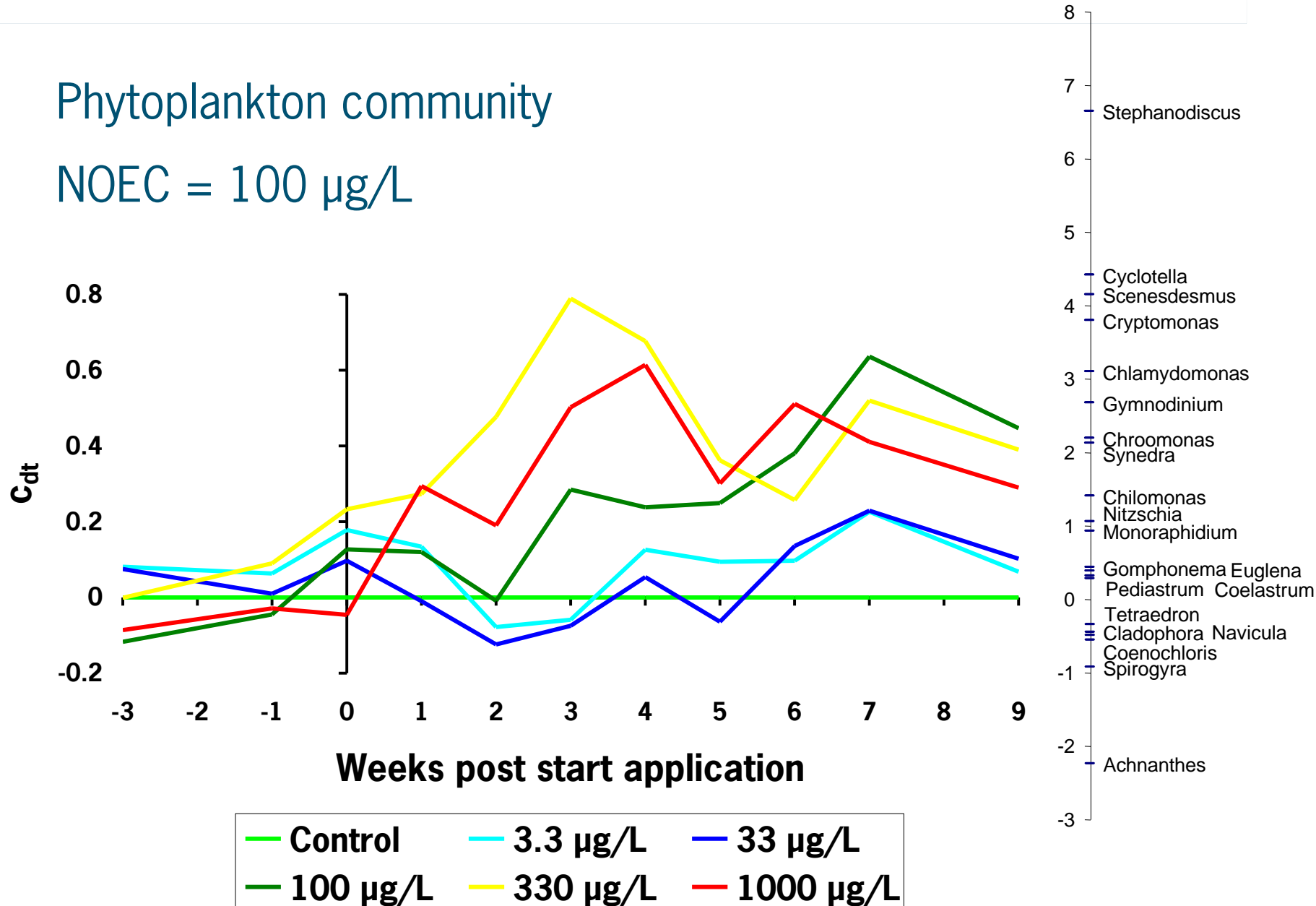


Lecane sp (NOEC = 330 µg/L) Indirect effect

Carbendazim microcosm experiment

Phytoplankton community

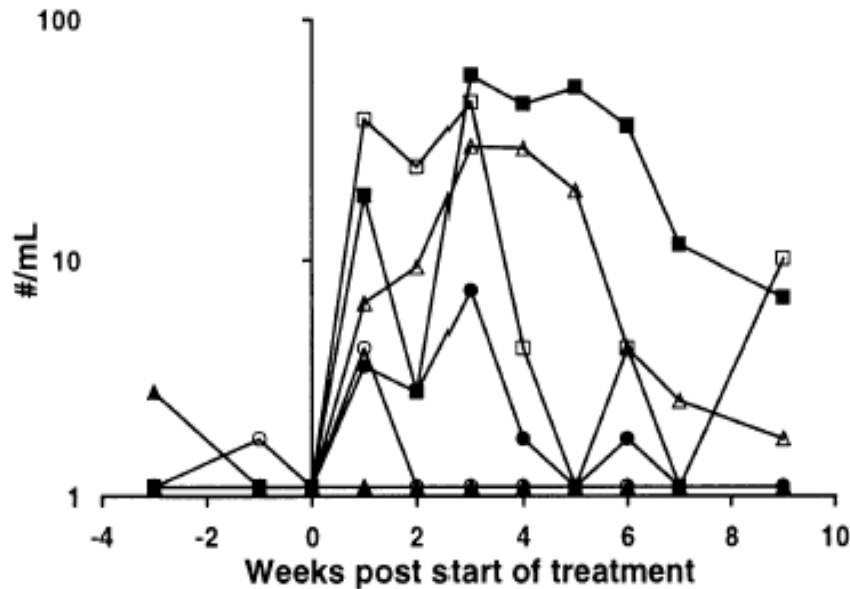
NOEC = 100 $\mu\text{g/L}$



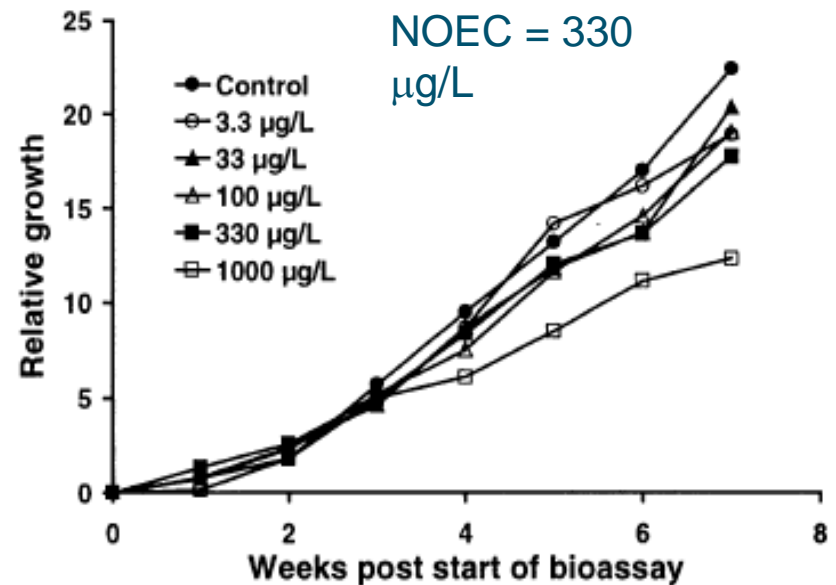
(In)direct effects carbendazim on *Scenedesmus* sp.



Free-living in microcosm



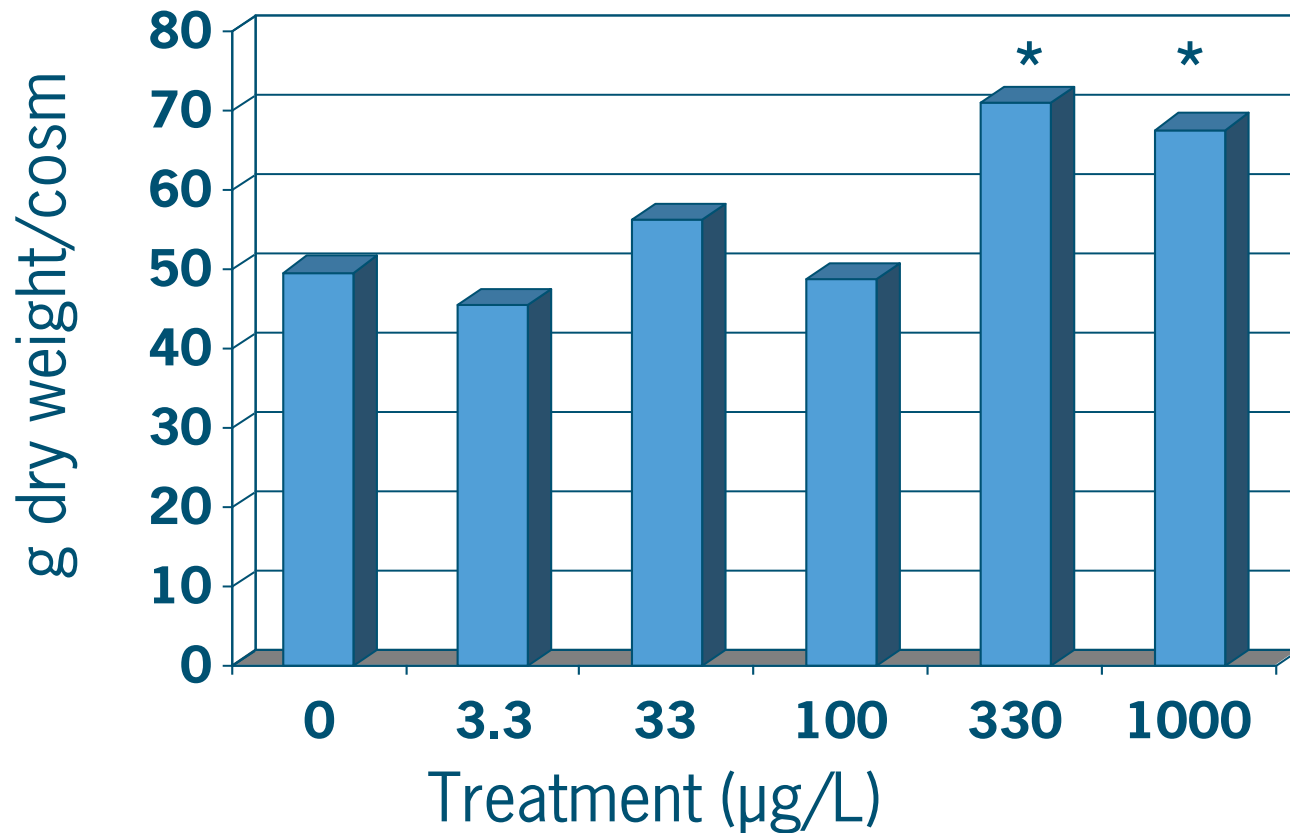
Bioassay



Influence decline of sensitive grazers may be larger than stress by toxicant

Carbendazim microcosm experiment

Dry weight biomass of *Elodea nuttallii* at end of experiment

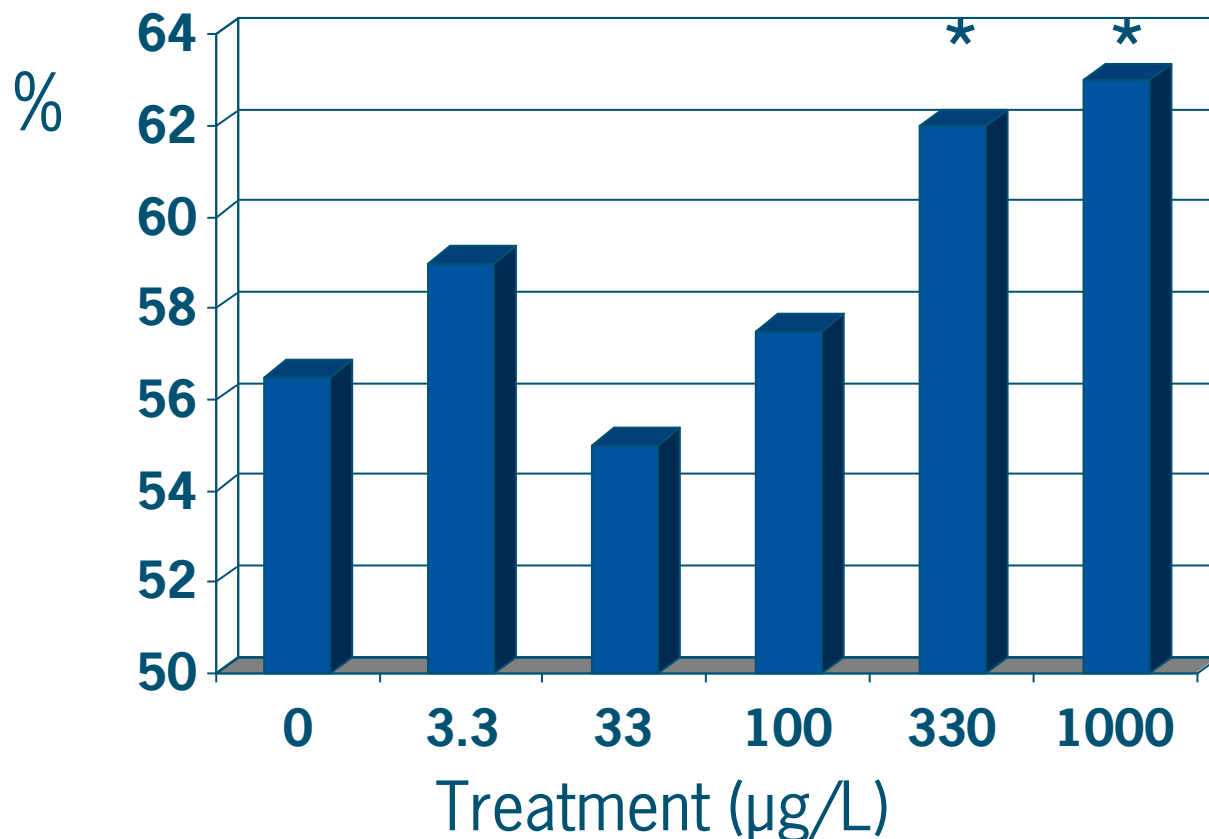


* $P < 0.05$



Carbendazim microcosm experiment

Residual dry weight of decomposing *Populus* leaves in small mesh litter bags after 4 weeks (exposure period)



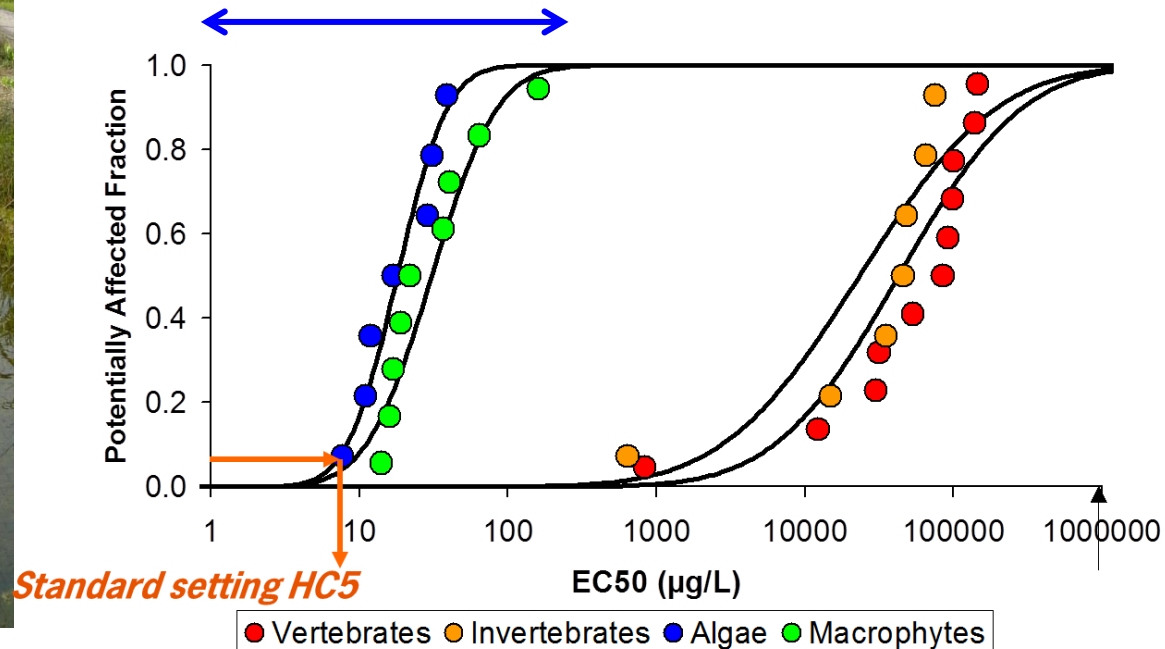
(In)direct effects herbicide metribuzin



- Field enclosure experiment
- single application metribuzin
- 0 – 1.8 – 5.6 – 18 – 56 – 180 $\mu\text{g/L}$
- dissipation DT50 = 6.5-9.4 day

SSD acute laboratory toxicity data

Concentration range enclosure experiment

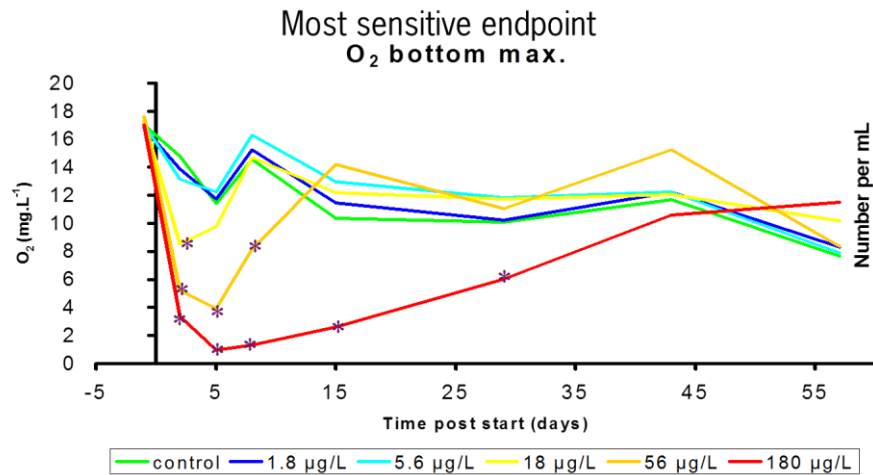


Laboratory toxicity data suggest that primary producers will suffer direct toxic effects at the exposure concentrations tested

(Brock et al. 2004: *Environ Poll* 130:403-426)

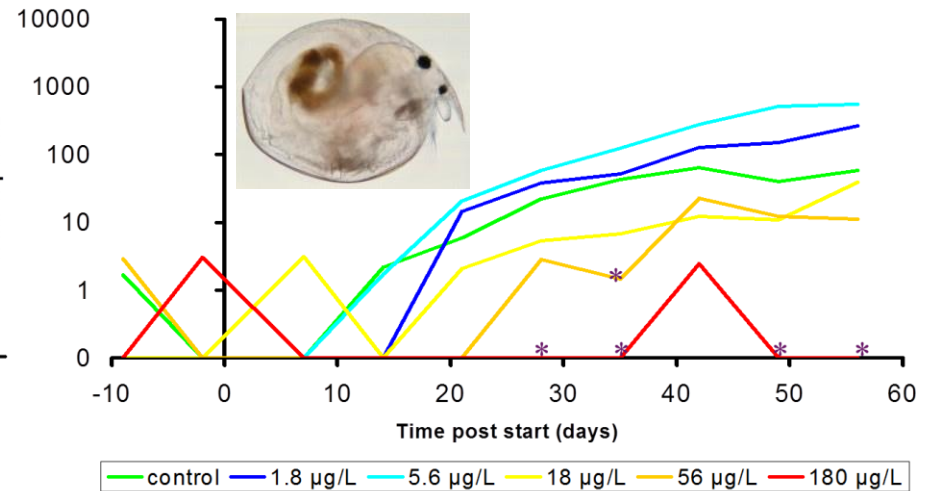
Indirect effects may persist longer than direct effects

Direct effects

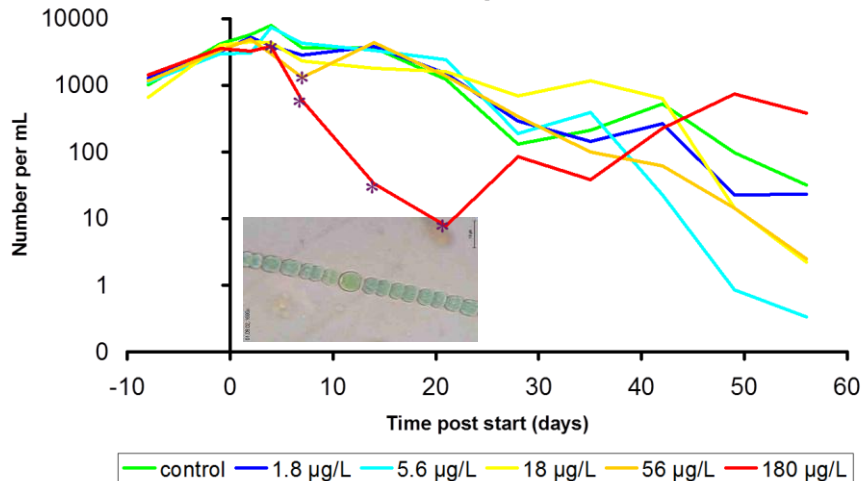


Indirect effects

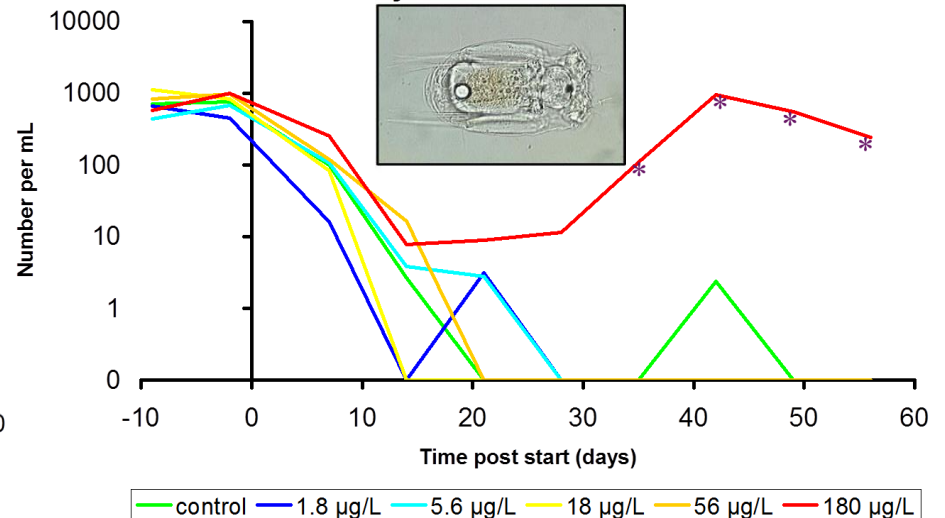
Chydorus sphaericus



Anabaena cf. cylindrica



Polyarthra remate



What type of ecological risk can be assessed in micro/mesocosms?

- Concentration-response relationships of dominant populations
 - Univariate statistics
- Concentration-response relationships of community
 - Multivariate techniques
- Relatively large species with complex life cycles are underrepresented in microcosms
 - Diversity of macrophytes is low
 - No large predators (e.g. fish)
- Isolated micro/mesocosms
 - recovery potential of r-selected species

Thank you for your attention

Questions ?



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